University Hospital Southampton	
NHS Foundation Trust	



	NHS	
ntral	& South	

Date:

 $\textbf{Specimen(s) received (central):} \ \texttt{BMT/Slides/BMAEDTA/PBEDTA/Cyto/other}$ MGG Flow Iron WGLS Sot. Flow WGLS Sal. WGLS Sot. Specimen(s) distribution: Histo

WGLS Sal. WGLS Sal.

паетнопс.	iest ne	quest	FUIIII		Specime	en(s) rece	eivea (ioca	aı):			вагсоае
Referrer Information	n				Patient II	nforma	ation				
Referring consultan	t				Surname						
Hospital					First Nam	ne					
Department					Date of B	Birth				Gender	
Departmental Emai	l (secure)				Hospital	numbe	er				
Telephone number	/ bleep				NHS num	ber					
□ NHS □ Pr	ivate				Address					Postcode:	:
Diagnosis / clinical How does the patie the National Genon Directory eligibility Clinical utility Priority	nt meet nic Test ?	Urgent □	anagement Cancer pathw			e□	Pleas	se notify the	laborato	sk? Yes □ ry if clinica	
Sample details (see	reverse for	instructio	ns)	Oper	ator:			Tel./B	leep		
Lab reference number				Colle					macro.	UHS lab	use only
(referrals to UHS)		ם סס		date/		ties 🗖	CCE		AB USE	tissus 🗆	Othor 🗆
Specimen type Refer to sample requirements overl Please indicate if multiples of the so collected eg. BM x2		PB □ BMA □ BMT □ Cytogenetics □ CSF □ FFPE □ Fresh tissue □ Other □ (EDTA) (EDTA) Biopsy □ Resection □ □ Trial (sent direct):									
	Haematology request(s) ✓ PB morphology □ BM morphology □ Iron stain □ BMT (haem) □ BMT □ CD138 IHC □ Congo Red □										
Flow cytometry tes	t request(s)	✓ One EDTA	per sub-section								
Acute Inc. blast enumeration	Acute □ Lymphoid □ Ext. B □ Ext. T □ Plasma cell □ Flow MRD □					BM / PB)					
Wessex Genomics I	_aboratory S	Service (So	uthampton) tes	st req	uest(s) ✓	One EDT	「A per su	b-section			
JAK2 V617F (monit	oring)		BRAF V600E					MYD88 L	265P		
Post allo transplant	chimerism ^b	5 🗆	FVL/PTM gend	otypin	g			CLL <i>IGHV</i>	somatic h	nypermutat	tion \square
<i>BCR::ABL1</i> qPCR – F	Presentation ¹		BCR::ABL1 qP0	CR –	Monitoring	¹		BCR::ABL	1 kinase d	lomain muta	ation** \square
Whole Genome Sec	uencing**		Acute leukaemia	molec	ular (geneti	ics) MRI	D ² **	Clonality	1	T cell c	disease 🗆
Eligible tumour** p			AML Follow up	П (АМ	(AML baseline from WGLS Salisbury³) Follow up		ury³)	A progressive testing strategy is used B cell disease [
Paired Germline (ap		0-14) 🗆	ALL Baseline				A progressive testing strategy is used D CCII GISCOSC				
LyConfirm no circulating tumour cells Date of diagnosis/this timepoint: Must be confirmed before germline sample is collected. Please refer to central NHS England eligibility criteria. Ensure clinical details are entered. MRD Marker:			imenoint:			Include histology / immunophenotyping report					
			-								
			Transcript/mutation type:								
Wessex Genomics Laboratory Service (Salisbury) test request(s) ✓ (see reverse for sample details)											
Myeloid disorders (MDS, MPN, AML, SM, CML***) ALL (T- and B-)											
G-banding						G-banding FISH SNP array			ay 🗆		
MPN panel (JAK2 V617F/CALR/MPL/JAK2 exon 12) Lymphoid (Mature: CLL, NHL, etc.)					-						
KIT D816V (ddPCR) \square Ext. KIT NGS panel (if D816V neg) \square					FISH ☐ TP53 NGS seq. ☐						
FIP1L1-PDGFRA (diagnosis) Lymphoid NGS panel											
AML (urgent molecular testing)					Myelo			_			
FLT3-ITD FLT3-TKD NPM1 IDH1 TP53 ***CML chronic phase is not a clinical indication for myeloid NGS panel			_			8-positive separation only (storage)					
Myeloma FISH panel											
Requester											
Name			Sign	ature						Di	ate

Email / phone

Sample transport

The referring laboratory is responsible for the safe transfer of tissue and it is thus recommended that Royal Mail Recorded Delivery or an equivalent tracked postal service is used.

Transport tracking ID

Please send flow samples to:	WGLS (Southampton) samples to:	WGLS (Salisbury) samples to:	BMT samples to:
Department of Immunology Laboratory Medicine University Hospital Southampton NHFT, Tremona Road, Southampton, Hampshire, SO16 6YD.	Formerly Department of Molecular Pathology Duthie Link Building, Mailpoint 225, University Hospital Southampton NHSFT, Tremona Road, Southampton, Hampshire, SO16 6YD.	Formerly Wessex Regional Genetics Laboratory Salisbury District Hospital, Salisbury, Wiltshire, SP2 8BJ	Department of Cellular Pathology, Pathology Block, Level E, Mailpoint 002. University Hospital Southampton NHSFT, Tremona Road, Southampton, Hampshire, SO16 6YD.
UKAS ref: 8696 20381 206615 / 02381 206640	UKAS ref: 9194 ☎ 02381 206638	UKAS ref: 1175 ☎ 01722 429080	UKAS Ref: 8178 ☎ 023 8120 4879
immunologylab@uhs.nhs.uk	wgls_cancergenomics@uhs.nhs.uk	shc-tr.WRGLdutyscientist@nhs.net	cellpath@uhs.nhs.uk
www.uhs.nhs.uk/health-professionals/pathology- services/handbook/laboratorymedicine	■ wgs_cancerwgls@uhs.nhs.uk (wgs only)	www.uhs.nhs.uk/health-professionals/pathology- services/handbook/cellularpathology	

Sample requirements

Details on both the referral card and the sample tube should be complete and legible. A minimum of 3 ID points are required. We reserve the right to refuse to process samples with incomplete, illegible or ambiguous patient information. Any samples in the wrong tube or medium, or which are subject to significant delay in transit, are liable to be rejected.

- a. FOR ALL GENOMIC TESTING, PLEASE ALSO PROVIDE SPECIFIC INFORMATION DETAILING HOW THE PATIENT MEETS THE NATIONAL GENOMIC TEST DIRECTORY (NGTD) ELIGIBILITY CRITERIA FOR THE TEST BEING REQUESTED (see http://www.england.nhs.uk/publication/national-genomic-test-directories for further information)
- b. FOR ALL GENOMIC TESTING, PLEASE INDICATE HOW TESTING WILL IMPACT PATIENT CARE. PATIENT MANAGEMENT INCLUDES DETERMINING THERAPEUTIC DECISIONS AND/OR CLINICAL INVESTIGATION AND/OR SURVEILLANCE PROGRAMMES.

Flow cytometry EDTA whole blood, bone marrow or fluid in universal container.

Sample must arrive and be tested within 72 hours of collection

cyte

WGLS (Southampton)
Formerly Molecular Pathology

FFPE: Send FFPE block (preferred) or tissue scrolls (3 x 20μm). Scrolls should be prepared on a clean microtome, ideally using a fresh blade per case, to avoid the risk of cross-contamination.

PB: Send minimum 3.4ml EDTA as an independent sample. This must **not** have been used previously on an automated laboratory analyser. ¹For *BCR::ABL1* analysis, if PB sample, please send minimum 12ml EDTA. TKI information and dates must be provided in clinical details to enable effective monitoring.

- ² For MRD analysis, if PB, please send 20ml EDTA, if BM, please send 5ml EDTA. Patients monitored at unusual laboratories must be highlighted and the MRD lab identified.
- ³ AML cases where the diagnostic sample identifies an MRD marker as part of the usual genetic work up, will have material sent from the WGLS Salisbury sample (the standard cytogenetics sample <u>no additional sample required specifically for AML baseline MRD</u>) to WGLS Southampton for RNA extraction and onward sending to the relevant laboratory.
- ⁴Adult patients (>26years of age), TYA patients (16-25years of age) only. Paediatric patients remain under the care of the CNS/trials team(s). Samples for RNA extraction must arrive in this laboratory within 48 hours of collection.
- ⁵ This laboratory performs whole PB/BM chimerism. This is not requesting Lineage Specific Chimerism, which is sent directly from clinical teams. ** This is a send away test.

BM:

- Conventional cytogenetics for acute leukaemias, MDS, MPD and AA: 0.5- 1ml in transport medium (but lithium heparin accepted)
- For new paediatric acute leukaemias also send KCH (3 drops of BM in KCH to be fixed at referring laboratory); if transport medium is not available, send in lithium heparin.
- Molecular studies of FLT3-ITD, FLT3-TKD, NPM1, IDH1, IDH2, TP53, KIT, JAK2, JAK2 exon 12, CALR, MPL and Myeloid NGS panel: 2-3ml in EDTA; however, material sent in BM transport medium or lithium heparin is acceptable.

PB:

- Conventional cytogenetics for new diagnosis CML, myelofibrosis or new acute leukaemias if no BM available: 5ml in lithium heparin.
- FISH for CLL/MCL and *TP53* mutation testing: 5ml in lithium heparin.
- Molecular studies of JAK2, CALR, MPL, myeloid NGS panel, etc.: 5ml in EDTA.
- FIP1L1-PDGFRA: 10ml in EDTA.

Smears: for CLL or NHL with suitable FISH markers, FISH can be attempted on freshly made, unfixed, unstained smears (at least 4 smears) if no fresh material is available. Please note that FISH can be attempted on smears that have been stored for some time if no other material available. Biopsies: FFPE sections / tumour dabs: slides containing unstained FFPE sections (3-4µm) or tumour dabs should contain at least 2 patient identifiers. Please package in a slide box. For large FFPE sections, please also send H&E slide with the tumour area appropriately marked. Other Tissues: lymph node, spleen, skin etc. should be sent in transport medium upon previous discussion with the laboratory. Please phone as soon as possible if anything is sent by courier which might arrive outside normal working hours. There is an on-call rota for acute presentations and Burkitt lymphoma after hours on a Friday or at the weekend (please phone the switch board on 01722 336262). In submitting samples the clinician confirms that consent has been obtained for testing and storage. Anonymised stored samples may be used for quality control procedures including validation of new genetic tests.

Cellular Pathology Specimen to be submitted in 60ml pot containing 10% neutral buffered formalin (ratio of 10:1). Direct purchase from Genta Medical.